

## QS S Assist **KINASE**\_MSA Kit

### Description

**KINASE** MSA Kit is designed for use in pharmacological assays for **KINASE** based on Off-chip mobility shift assay (MSA). This kit includes Assay Buffer, Termination Buffer, human protein kinase, ATP/Substrate/Metal and a protocol to perform 384 well plate assay.

### Components (400dpt x 1set)

| Materials               | Volume           | Storage                 |
|-------------------------|------------------|-------------------------|
| 10 x Assay Buffer       | 5 mL x 1         | -80°C                   |
| <b>KINASE</b> *         | <b>25</b> µL x 1 | -80°C                   |
| 5 x ATP/substrate/metal | 0.6 mL x 1       | -80°C (light shielding) |
| 4 x Termination Buffer  | 7.5 mL x 1       | -80°C                   |

\* Concentration of the kinase; **100** µg/mL  
Please avoid repeated freeze-thaw cycles.

### Reagent Preparation (per 400 dpt)

Bring all reagents (except kinases) to room temperature before use.

### Materials provided

#### **Assay Buffer**

Thaw 10 x Assay Buffer and take 4 mL. Dilute with 36 mL of distilled water (total volume: 40 mL). Adjusted Assay Buffer is able to keep room temperature before use. Please do not carry over this buffer on the next day, because the buffer component DTT is unstable.

#### **ATP/Substrate/Metal Solution**

Thaw 5 x ATP/Substrate/Metal component and dilute 0.45 mL into 1.8 mL of distilled water (total volume: 2.25 mL). Bring the solution to room temperature and shield from light until use.

#### **Enzyme Solution**

Thaw **KINASE** and dilute it appropriate-fold with Assay Buffer. Please keep it on ice before use.

#### **Termination Buffer**

Thaw 4 x Termination Buffer and dilute 7 mL into 21 mL of distilled water (total volume: 28 mL). Bring Termination Buffer to room temperature ( $25 \pm 2^\circ\text{C}$ ) prior to use.

## Materials required

### Compound Solution

Prepare a hundred times concentrated compound stock solution with DMSO. Dilute the solution 25 times with Assay Buffer. For the vehicle control, prepare 4% DMSO-Assay Buffer solution.

**Coating-3 Reagent (CR-3) 30 mL bottle from PerkinElmer Inc., cat. no. 760050**

### Separation Buffer

100 mM HEPES (pH7.5), 10 mM MgCl<sub>2</sub>, 0.01 % Triton X-100, 10 mM EDTA-2Na,  
1 % DMSO, 0.1 % Coating-3 Reagent.

### Example of Reaction Mixture

| Sample | Compound Solution<br>( $\mu$ L) | Vehicle<br>( $\mu$ L) | ATP/Substrate/Metal<br>Solution<br>( $\mu$ L) | Enzyme<br>Solution<br>( $\mu$ L) | Assay Buffer<br>( $\mu$ L) |
|--------|---------------------------------|-----------------------|---|----------------------------------|----------------------------|
| A      | —                               | 5                     | 5   | —                                | 10                         |
| B      | —                               | 5                     | 5   | 10                               | —                          |
| C      | 5                               | —                     | 5   | 10                               | —                          |

Where A equals negative control, B equals positive control and C equals test sample.

Calculate the percent inhibition of compound as follows;

$$\text{Inhibition (\%)} = (1 - (C - A) / (B - A)) \times 100$$

### Final Concentrations of Components in Reaction Mixture

20 mM HEPES (pH7.5), 0.01 % Triton X-100, 2 mM DTT

1000 nM Substrate, 10  $\mu$ M ATP, 5 mM Metal

### ASSAY PROCEDURE:

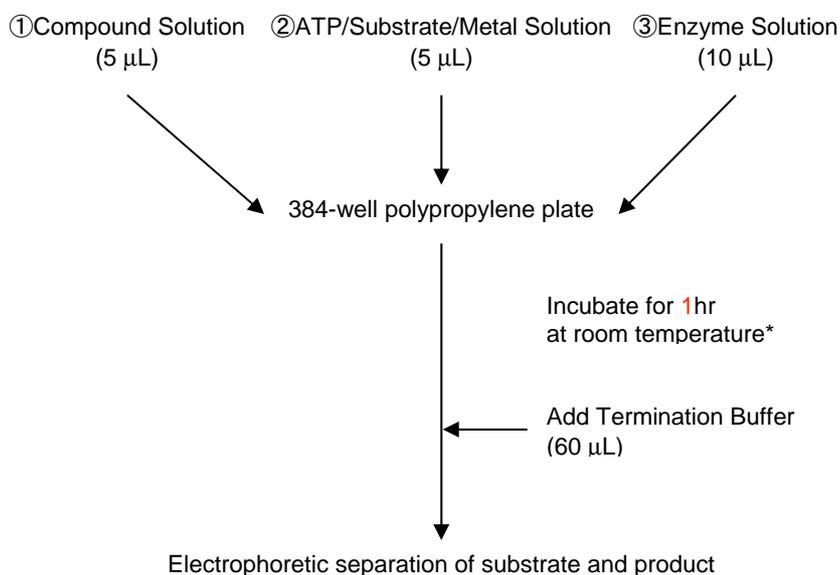
All procedures are performed at room temperature.

1. Add 5  $\mu$ L of vehicle (4% DMSO) to wells of “A” and “B” and Compound Solution to well of “C” of a 384-well assay plate.
2. Add 5  $\mu$ L of ATP/Substrate/Metal Solution to each well.
3. Add 10  $\mu$ L of Assay Buffer to wells of “A” and Enzyme Solution to wells of “B” and “C” to start kinase reaction. Cover the plate and incubate for **1** hour at room temperature.
4. Add 60  $\mu$ L of Termination Buffer to stop kinase reaction.

Place plate(s) into MSA device.

5. Measure the Product/Sum ratio.

### Illustration of Assay Procedures:



\*Depend on Enzyme

**Separation conditions for 12-Sipper chip**

| Parameter                         | Setting  |
|-----------------------------------|--|
| Chip                              | 12-Sipper Chip   |
| Analysis Results Type             | Off-Chip Mobility Shift  |
| Threshold                         | None   |
| Baseline Threshold                | None   |
| Peak Order                        | <b>Product</b> First   |
| Downstream (V)                    | - 600  |
| Upstream (V)                      | - 1800   |
| Base Pressure (psi)               | - 0.1 (760404) / - 0.5 (760394)<br>Note: Please adjust base pressure (psi) to the chip type you apply. |
| Pressure (psi)                    | - 1.1  |
| Initial Delay (sec)               | 0  |
| End of Plate Delay (sec)          | 70   |
| Final Delay (sec)                 | 17   |
| Sample sip Time (sec)             | 0.2  |
| Post Sample Buffer sip Time (sec) | 35   |
| Dye sip Time (sec)                | 0.2  |
| Post Dye sip Time (sec)           | 45   |

**Detection System**

|              |              |
|--------------|--------------|
| Light Source | Blue Laser   |
| Excitation   | 488 nm       |
| Emission     | 510 - 550 nm |

The typical separation pattern of substrate and product with **KINASE MSA Kit** is shown below.

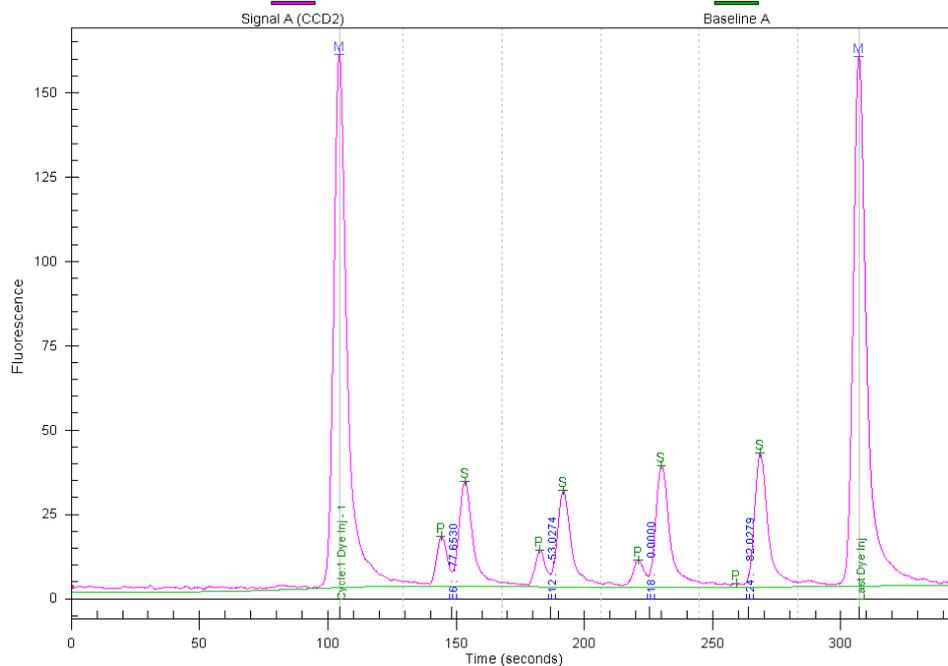


Fig.1 Typical separation pattern under the described separation conditions on page 4.

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