

Activity-Based Kinase Selectivity Profiling of Clinically Relevant Tyrosine Kinase Inhibitors using QuickScout™

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Introduction

Protein kinases are crucial elements of inter- and intra-cellular signaling and, as such, have been important targets for the development of activity-modulating effectors. To date, the most relevant clinical application of using kinase inhibitors to disrupt cell signaling pathways has been in the area of oncology. In recent years, eight distinct tyrosine kinase inhibitors have been approved globally for adjunctive or primary cancer therapy. These molecular inhibitors were rationally designed to target specific tyrosine kinases, and most bind competitively, at or near the ATP pocket. Because of the competitive nature of inhibitor binding, changes in the concentration of ATP present in any such reaction may lead to unanticipated results. Here we describe the use of our QuickScout™ activity-based-kinase assay system to determine the full selectivity profiles of the eight commercially available, clinically-approved protein kinase inhibitors against our panel of 79 tyrosine, 197 serine/threonine, 24 mutant and 3 lipid kinases. In this study, all assays were performed at two different ATP concentrations - near the K_m^{ATP} and at 1 mM ATP. This comparative analysis should provide useful insight for the development of additional protein kinase inhibitors as therapeutic agents.

Materials and Methods

All human recombinant protein kinases were made in Carina Biosciences, Inc. (Kobe, Japan) as indicated in its web site (<http://www.carnabio.com>). All kinase assays and profiling (% Inhibition and IC₅₀ Determination) of each drug were performed according to the procedures indicated in the web site of Carina Biosciences, Inc. The kinase activities were determined by off-chip mobility shift assay using LabChip™3000 (Caliper LS) and fluorescence polarization assay using IMAP™ technology or ELISA. Clinically relevant tyrosine kinase inhibitors (right table) were used for the present experiments.

Results

Each of the eight approved kinase inhibitors were screened at a concentration of 1 μM against the QuickScout™ panel of >300 human kinases, to identify candidate kinase targets (Primary Screen). Quantitative IC₅₀ determinations were then made for each kinase-inhibitor against each kinase identified in the primary screen. Where inhibition >40% was observed in these screens, additional testing was performed using approved inhibitors (and staurosporine, as positive control) at measured IC₅₀ concentrations, and ATP at both K_m^{ATP} and 1 mM (physiological) concentrations. 'Kinome Clustering' results of these experiments are shown graphically in Figure 1. In Figure 2, we show a novel approach for the visualization of inhibitor selectivity by ranking IC₅₀ values obtained using physiological (1 mM) ATP levels. Inhibitor selectivity was further analyzed using published pharmacokinetic parameters describing plasma concentration values, C_{min} . These results are shown in Figure 3.

In Table 1, we show data comparing the results of our 'physiological' assays (performed using 1 mM ATP) with reported results from cellular proliferation assays. Finally, we report our inhibitory activity results as they relate to 'on-target' kinases and clinically significant mutant forms (Table 2).

Kinome Tree

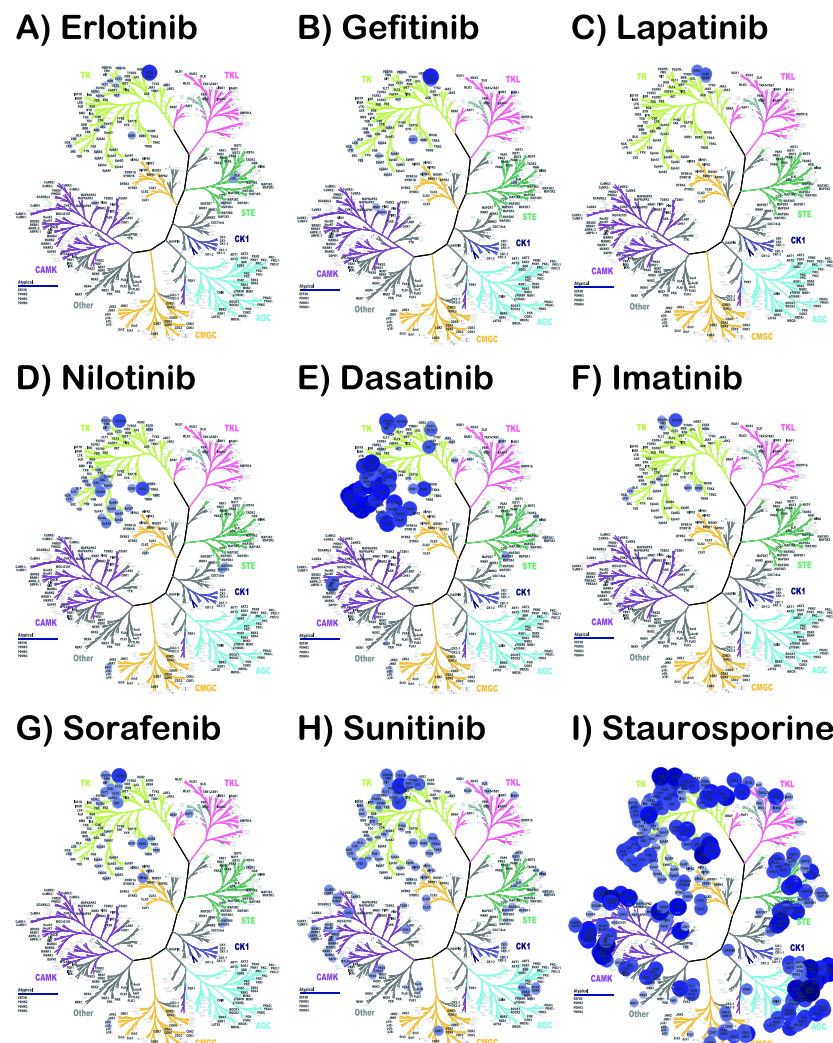
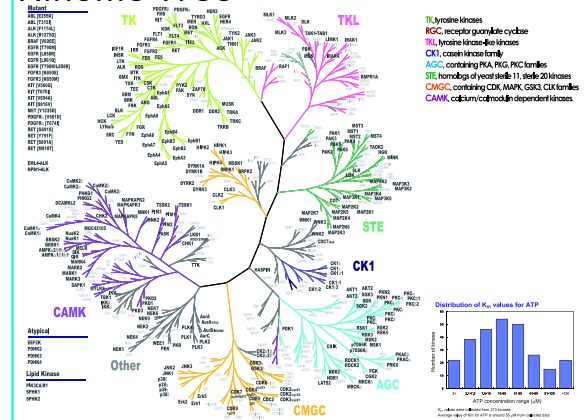


Figure 1

QuickScout™ profiling (ATP conc. = K_m^{ATP}) results of approved 8 kinase inhibitors and staurosporine. Dendrograms were adapted to human kinome tree. Kinase inhibition maps for inhibitors, A) erlotinib, B) gefitinib C) lapatinib D) nilotinib E) dasatinib F) imatinib G) sorafenib H) sunitinib and I) staurosporine, respectively. Inhibition concentration (IC₅₀) values under 10,000 nM are marked with blue circles where larger circles indicated strongly inhibit to kinase.

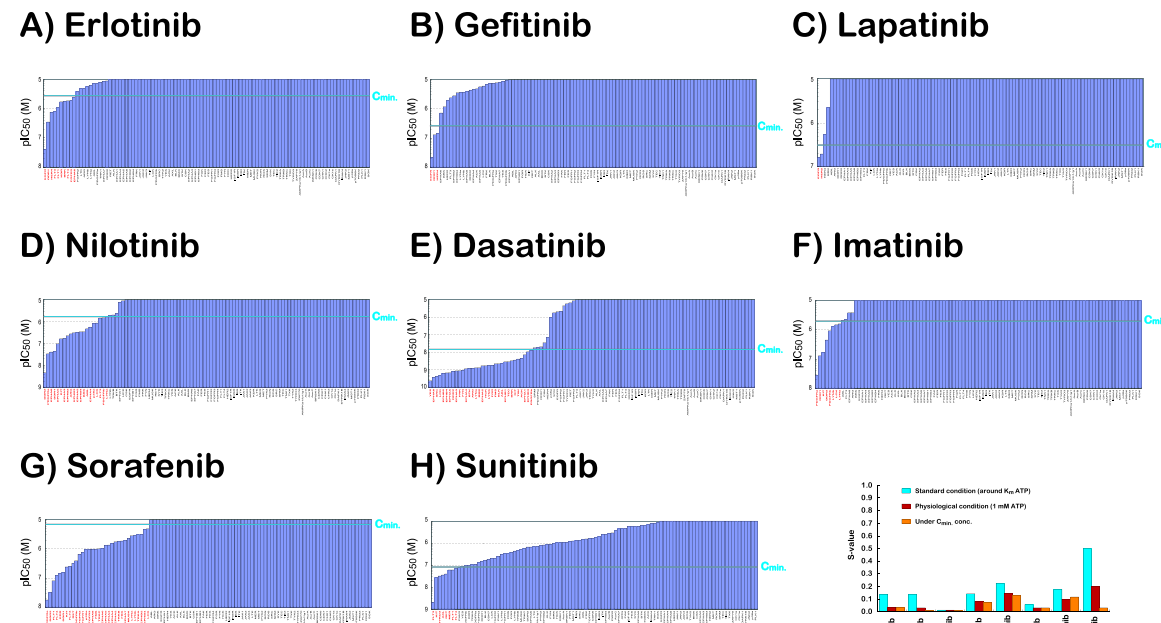


Figure 2

QuickScout™ profiling (ATP conc. = 1 mM) results of approved 8 kinase inhibitors. Top hit 100 kinases were selected from results in figure 1 and IC₅₀ values were determined using 1 mM ATP. Kinases were aligned in ascending order of the IC₅₀ values. Mutant kinases were excluded from this analysis. The blue line indicates the C_{min} plasma concentration (mol/L) of the drugs in patients. These C_{min} concentrations were reported in Cancer Treatment Reviews (2009) 35: 692-706 (erlotinib, gefitinib, lapatinib, nilotinib, imatinib, and sunitinib), Journal of Clinical Oncology, (2008) 26: ASCO abstract #3590. (dasatinib) and Clinical Cancer Res. (2006) 12: 144-151 (sorafenib).

Table 2

IC₅₀ effective ratios at 1mM ATP condition assay Wild type kinase vs Mutants

EGFR	Erlotinib IC ₅₀ (nM)	Ratio	Gefitinib IC ₅₀ (nM)	Ratio	Lapatinib IC ₅₀ (nM)	Ratio
W.T.	39.8	1.0	22.0	1.0	164.0	1.0
T790M	>10000	N.D.	>10000	N.D.	>10000	N.D.
T790M/L858R	>10000	N.D.	>10000	N.D.	>10000	N.D.
L858R	9.6	4.1	6.0	3.6	144.4	1.1
L861Q	27.7	1.4	12.5	1.8	52.2	3.1

ABL	Nilotinib IC ₅₀ (nM)	Ratio	Dasatinib IC ₅₀ (nM)	Ratio	Imatinib IC ₅₀ (nM)	Ratio
W.T.	471.4	1.0	2.3	1.0	2144.6	1.0
E255K	>10000	N.D.	10.0	0.2	>10000	N.D.
T315I	>10000	N.D.	>10000	N.D.	>10000	N.D.

KIT	Nilotinib IC ₅₀ (nM)	Ratio	Dasatinib IC ₅₀ (nM)	Ratio	Imatinib IC ₅₀ (nM)	Ratio
W.T.	166.4	1.0	3.7	1.0	168.1	1.0
V560G	6.0	28	0.4	9.3	9.5	18
V564A	>10000	N.D.	37.6	0.1	>10000	N.D.
T670I	7332.3	0.02	>10000	N.D.	>10000	N.D.
D816V	>10000	N.D.	55.4	0.07	>10000	N.D.

PDGFRα	Nilotinib IC ₅₀ (nM)	Ratio	Dasatinib IC ₅₀ (nM)	Ratio	Imatinib IC ₅₀ (nM)	Ratio
W.T.	34.6	1.0	19.3	1.0	26.8	1.0
V561D	201.6	0.17	26.0	0.17	87.7	0.33
T674I	>10000	N.D.	>10000	N.D.	>10000	N.D.

RET	Sunitinib IC ₅₀ (nM)	Ratio	Sorafenib IC ₅₀ (nM)	Ratio
W.T.	322.5	1.0	39.1	1.0
G691S	448.3	0.72	50.3	0.78
Y791F	407.8	0.79	62.9	0.62
S891A	239.5	1.4	50.5	0.77
M918T	833.1	0.39	95.2	0.41

FGFR3	Sunitinib IC ₅₀ (nM)	Ratio	Sorafenib IC ₅₀ (nM)	Ratio
W.T.	974.6	1.0	128.6	1.0
K650E	725.8	1.3	264.1	0.49
K650M	1250.1	0.78	540.8	0.24

Figure 3

Selectivity as quantitative measure of specificity. Selectivity (S-values) calculated for kinases with IC₅₀ values < 3 μM at both conditions of an around ATP Km (Standard, Blue bars) and 1 mM ATP (physiological, Red bars) with following equation: Number of kinases (IC₅₀ values < 3 μM) / tested kinase. Orange bars indicate that S-values calculated from count of kinase in under C_{min} concentration per tested kinase, with following equation: Number of kinases (IC₅₀ values < C_{min}) / tested kinase. Mutant kinases were excluded from this analysis.

Conclusion

- Selectivity profiling was performed for each of the eight clinically approved kinase inhibitors against a panel containing 79 tyrosine-, 197 serine/threonine-, and 3 lipid kinases at ATP concentrations of K_m^{ATP} , and 1 mM.
- Under standard profiling conditions (ATP concentration approximately Km), the Selectivity Ranking (number of kinases with IC₅₀ < 3 μM / number of kinases tested) was: lapatinib < imatinib < erlotinib = gefitinib < nilotinib < sorafenib < dasatinib < sunitinib.
- Under physiological profiling conditions (ATP concentration = 1 mM), the Selectivity Ranking (number of kinases with IC₅₀ < 3 μM / number of kinases tested) was: lapatinib < gefitinib < imatinib < erlotinib < nilotinib < sorafenib < dasatinib < sunitinib.
- When analysis incorporated the use of reported pharmacokinetic data with data from physiological profiling conditions (ATP concentration = 1 mM), the Target Selectivity Index (number of kinases with IC₅₀ values < plasma concentration / number of kinases tested) was: lapatinib = gefitinib < imatinib < sunitinib < erlotinib < nilotinib < dasatinib = sorafenib.
- Biochemical assays performed using 1 mM ATP resulted in IC₅₀ values very similar to those reported in Cellular Proliferation Assays for the approved kinase inhibitors.
- When tested against mutated target kinases at 1 mM ATP, all approved inhibitors showed less potency against gate-keeper mutants.
- Comparative selectivity profiling performed at both Km and 1 mM concentrations of ATP provide useful insight for the assessment of on-target and off-target kinase inhibitors.